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The Dataset Assembly Tool is a resource to make data wrangling faster and easier, to improve the quality and timeliness of IDI research.

This presentation includes additional details in the speaker's notes – reading through the presentation is also recommended.

You may want to print the presentation as "Notes Pages", with the slides and the presenter notes.

This presentation should be read/watched alongside "Accelerating Dataset Assembly: Primer and guide to the Dataset Assembly Tool", which discusses the underlying approach and provides further technical information as well as a worked example.

We include images of code. ACTUAL CODE IN THE TOOL MAY HAVE BEEN UPDATED SINCE THIS PRESENTATION WAS MADE. So notes refer to the code in the images in this presentation, users will have to use their best judgement when comparing this presentation to the working tool.

# SOCIAL WELLBEING WELLBEING TANGATA

Making dataset assembly easier

Data rich, structure poor

Rich and detailed data

data preparation

Many different datasets in the IDI

Without consistent format or logic

More datasets, more preparation

IDI projects can spend a long time on

IDI is a rich resource for research. But because the data is gathered for administrative purposes it is not well prepared for research use. Hence significant effort can be required to construct a suitable dataset for research – or researchers limit themselves to only a small part of the IDI.

Dataset assembly tool

analytics sooner

Test ideas faster

Goal to reduce preparation time

Move from data wrangling to

Create your own definitions

Share and reuse definitions

This is not the first tool developed to make it easier to pull different datasets together. However, where previous tools have seen limited reuse across different projects, the assembly tool has already been applied to multiple projects so we are confident that it offers something of wider value.

The tool was designed within a broader set of practices and patterns. When used in the context of these practices and patterns, it also supports knowledge sharing and provides further acceleration to projects via the reuse of materials between researchers and between projects. If researchers follow similar processes, then components of our research will be more easily shared, reused, and cited by our colleagues. This is discussed in further detail in "Accelerating Dataset Assembly: Primer and guide to the Dataset Assembly Tool".

### Why should I use this tool?

#### As an IDI researcher, I want to...

- Construct analysis ready, rectangular datasets from a range of data sources with ease
- 2. Keep different project stages separate and independent from each other
- 3. Design new variables without being required to fit a specific format
- 4. Add new variables to an existing project dataset with ease
- 5. Transfer definitions between projects as easy as copy & paste
- 6. Trace where data has originated from

- Replicate parts of the automated assembly manually for validation and guality assurance
- 8. Use and understand the tool without learning a new programming language
- 9. Debug only a small part of the process when an error occurs, and know rapidly which part is at fault

## If you want the same, this tool is for you



I assessed several existing tools (including my previous attempts) and identified these nine things I want to do as a researcher based on what needs these tools met, and where there was still frustration. The Dataset Assembly Tool was then built to address each of these needs.

How does the tool meet these needs?

- 1. Accepts lists of inputs, and outputs rectangular dataset
- 2. Definitions must run before assembly, most efficient to re-run assembly when adding new definitions than appending them on
- 3. Tool input adapts to different formats
- 4. Keeping assembly fast so re-assembling is easy
- 5. Definitions are separate from assembly
- 6. Input control files act as record of what was assembled & from where
- 7. Assembly tool uses a single core query query is saved to disc for inspection, and it easy to duplicate
- 8. Tool written in R, but control files are Excel (everyone can use Excel)
- 9. Multiple places of error checks with sensible messages

### Overview of how it works

### Approximate sequence

#### 1) IDI access

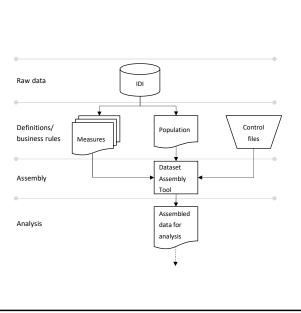
2a) Study population is defined

2b) Measure(s) of interest defined

3) Control files filled in with population and measure details

4) Dataset Assembly Tool executed

5) Analysis on rectangular dataset





- 1. Access to the raw data/potential
- 2. Researchers define their study population and the events/measures that are of interest to their research.
  - All of these definitions are independent from each other.
  - · Definitions could be copied in from other projects
  - No way to avoid having to define what you are interested in
- 3. Excel control files are populated
  - Includes where to get the definition from
  - How to summarise/prepare it
  - Control files becomes your initial data dictionary
- 4. Dataset Assembly Tool is executed
  - Control files are read & checked
  - Every measure is created for every population
- 5. Assembled data is available for research
  - (start with data cleaning e.g. how should missing values be handled)

More details and a worked example can be found in "Accelerating Dataset Assembly: Primer and guide to the Dataset Assembly Tool". Also the assembly tool code contains an example using IRD data.

### Who-when-what pattern of assembly

### Inputs

Population definition:

- Identities
- Start dates
- End dates

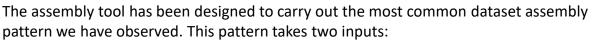
Events:

- Identity
- When it occurred
- Event details

### Outputs

For every identity

- A summary
- Of the events
- That occurred between start and end dates



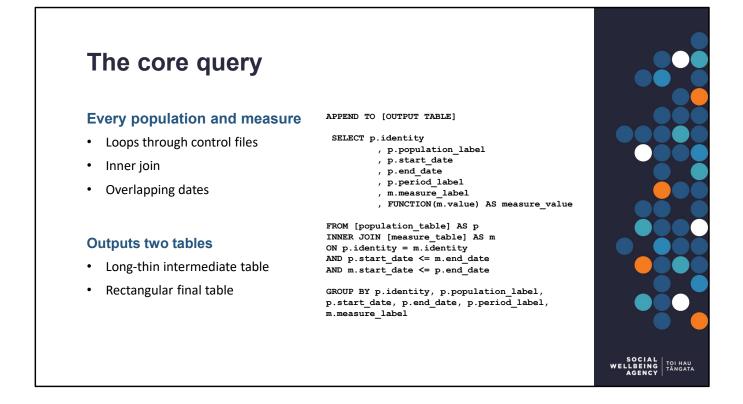
- 1. Population definition and a date range of interest
- 2. A (series of) event definitions with identities, dates and details

Defining each of these separately is usually straightforward.

### For example:

- 1. Population = A list of school leavers, from when they graduated until one-year later
- 2. Event = Income from wages & salaries reported to IRD
- These could be combined to output

For each school leaver, total earnings in the year following graduation.



One SQL query does all the main processing. This core query is shown on the right side. It is adapted for each row in the control file, but the overall pattern remains the same.

This core query outputs a long, thin table. This intermediate table is designed so it is easy to add all the measures to it using only the core query. This simplifies the tool, making it faster and less vulnerable to errors.

Once all the measures have been appended on to the intermediate table, the assembly tool pivots the table to create a rectangular final table that is ready for research use: One row per observation, and one column per measure.

Because the core query inner joins by identity and overlapping dates, researchers can define their research population completely separate from defining their measures. For my measures, I aim to create the best definition possible (for all people and time periods), knowing that the assembly tool will return only those details that apply to my study population and time period.



Before you can use the tool for the first time, some setup is required. This should only be necessary the first time.

Unless you follow these steps the tool will not work.

Skip over this section in a short presentation.

### Installation

### Copy tool into project

- Fetch from GitHub or IDI wiki
- Unzip
- Check contents are correct

#### Two setup tasks in two files

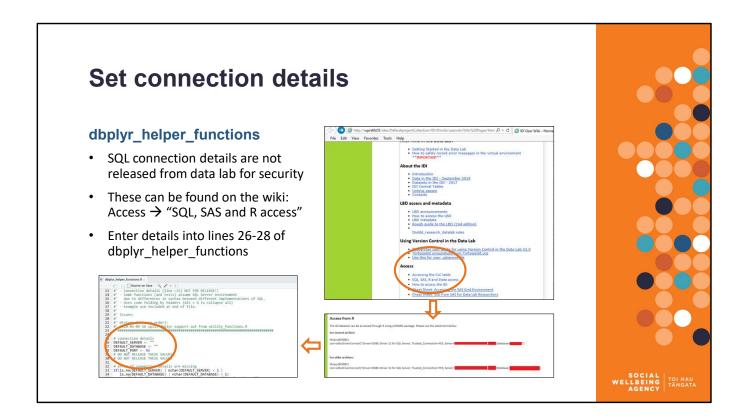
- dbplyr\_helper\_functions
- automated\_tests

Name	Date modified	Туре	Size
documentation	12/05/2020 9:22 AM	File folder	
tests	12/05/2020 9:23 AM	File folder	
automated_tests.R	8/05/2020 3:50 PM	R File	4 KB
dbplyr_helper_functions.R	12/05/2020 9:22 AM	R File	27 KB
general_assembly_tool.R	4/03/2020 4:45 PM	R File	20 KB
general_assembly_tool_functions.R	4/03/2020 4:43 PM	R File	5 KB
measures.xlsx	5/03/2020 9:58 AM	Microsoft Excel W	15 KB
population_and_period.xlsx	4/03/2020 3:06 PM	Microsoft Excel W	9 KB
run_assembly.R	18/03/2020 2:24 PM	R File	4 KB
table_consistency_checks.R	8/05/2020 4:22 PM	R File	12 KB
utility_functions.R	8/05/2020 4:12 PM	R File	7 KB

The tool is a set of R scripts, plus accompanying documentation, tests and exemplars. You can download these from SWA's GitHub page: https://github.com/nz-social-wellbeing-agency/dataset\_assembly\_tool Or check the IDI wiki for a copy that is already inside the datalab.

This should arrive bundled in a zip file. Copy into your IDI project and unzip. The image shows the expected contents of the tool.

Once you have unzipped the tool there are two tasks that need to be completed in two different files.



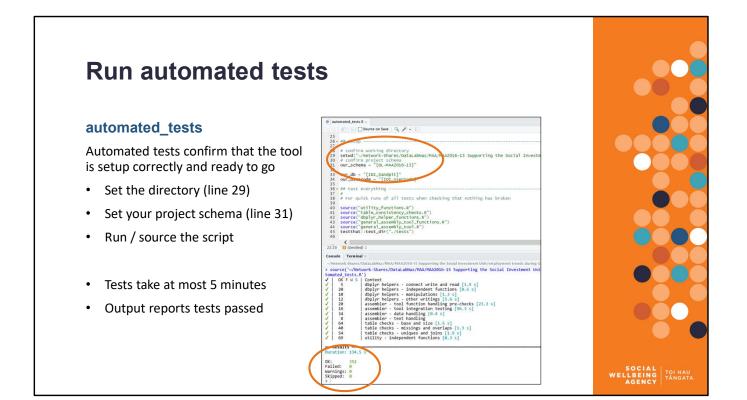
(Top left image) On the IDI wiki, under the heading "Access", click the link "SQ:, SAS, R and Stata access".

(Bottom right) Read off the server name, port number, and database under the "Access from R" heading

(Bottom left) Enter these values into the "connection details" section at the top of dbplyr\_helper\_functions.R

The final results looks like:

DEFAULT\_SERVER <- "[server.name.bond]" DEFAULT\_DATEBASE <- "[database.name.james]" DEFAULT\_PORT <- 007



The assembly tool is covered by unit record tests. These are tests that check all of the functions of the tool (and associated code) are performing correctly. They include copying a worked example onto the SQL server and checking that it returns the correct output.

All the tests and resources required to run them are found in the "tests" folder. However, all you need to access is "automated\_tests.R".

This script will run all the automated tests and output a summary. If all the tests pass then you have confirmed that the tool is setup and running correctly.

Top half of image is the testing script. The two lines that need to be updated are circled. Bottom half of image is the testing output:

- All the tests run
- Whether tests passed (OK), failed (F), gave warnings (W) or were skipped (S)
- Circled is the final summary showing all tests were passed.

The tests are best run in a new/empty session. If errors occur / the tests fail, then see the section at the end of this presentation of what to do when things go wrong.

### A note on packages

#### Tool works without installing packages

When Stats NZ updated the R Server in the IDI at the start of August 2020, they pre-installed a wide range of R packages.

The Dataset Assembly Tool works with all the pre-installed packages.

#### **Package versions**

- Over time some packages will need updating.
- Due to how packages interact this could cause the tool to fail.
- A list of installed packages from when the tool was last tested is provided.



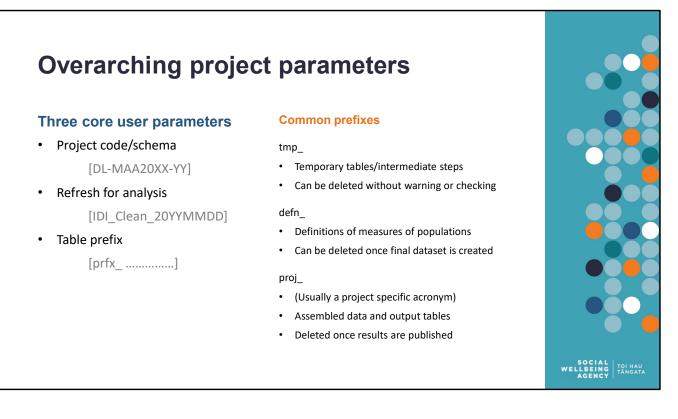
We also provide a list of the packages & versions installed with our working configuration of the tool. Some users with other versions of packages installed may experience errors. Ensuring that your package versions are consistent with ours reduces the risk of this. As Stats NZ has already installed a range of default packages, it should generally be sufficient to uninstall any version you have installed yourself and rely on the version Stats NZ provide to all users.

Package & version differences are more likely to cause warnings than errors. E.g. prior to August 2020 the tool would produce warnings from the 'rlang' package about some commands being depreciated. Warnings are unlikely to prevent the tool from running.



This section covers an example of using the tool. It demonstrates all the key workings that researchers will need.

As part of designing this example, I went from an empty project to a complete dataset in half a day.



Several parameters turn up in many places throughout a research project. Deciding these once at the start of the project helps ensure consistency throughout the project.

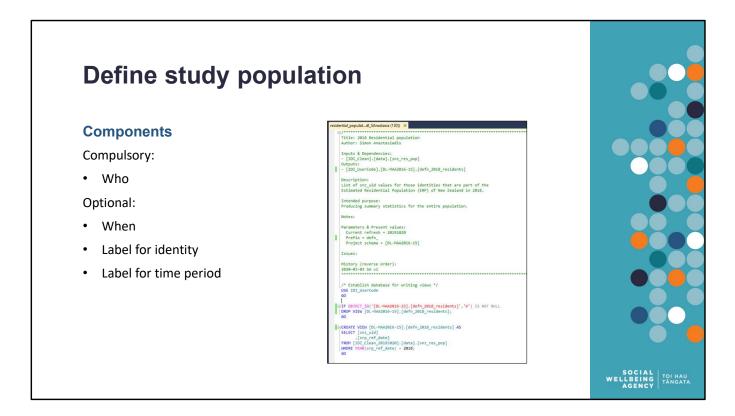
It is the user's responsibility to ensure that these parameters are consistent throughout their analysis.

(The tool does not check that you have used the same refresh or schema name throughout. The user is responsible for ensuring consistent parameters are used.)

A table prefix is optional, but highly recommended.

Because tables are sorted alphabetically in SQL Server, using a prefix keeps related tables together. It also makes purpose of table clear. Aids identification and removal of unneeded tables – being a good steward to data lab resources.

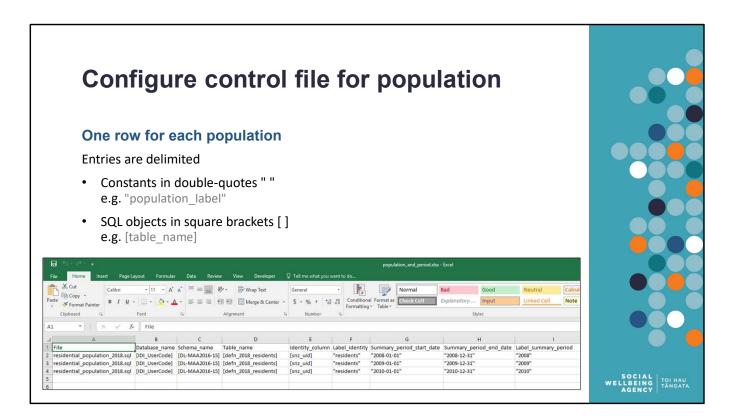
As an example of a project specific prefix, I used "chh\_" as the prefix for my analysis into "<u>c</u>onstructing <u>h</u>ouse<u>h</u>olds"



An SQL script must be written that defines your population of interest. It must contain 'who' is in the study population.

If when each person is being studies differs then the table should also contain when. If the same person is studied more than once (e.g. before and after groups), or in more than one time period, you may also wish to provide labels for the identity (or group of identities) or the time period.

More than one study population can be defined (E.g. one definition for treatment, and one definition for control), each in its own file.



Enter the details of each population in the table.

All columns other than File are compulsory. Every cell must be populated. Compulsory cells must be delimited.

Columns:

- Database, schema and table names give the SQL location where the table is saved
- Identity is the column that contains your individual identifiers (will most often be [snz\_uid] but could take other values, like meshblock codes if needed)
- Summary period start and end date define the period of time of interest for our population (Format: "YYYY-MM-DD", e.g. "2020-12-31")
- · Identity labels are assigned to the identities
- Summary period labels are assigned to the periods

The example control file, creates a panel of data for the same population for three years.

Details on each column of the population file

- 1. File (optional) Recommended you record the file name of the SQL script that creates your study population.
- 2. Database Name (required, sql object only) The name of the SQL database

where the study population is stored. Most likely [IDI\_Clean\_XXXX] or [IDI\_Sandpit] or [IDI\_UserCode].

- 3. Schema Name (required, sql object only) The name of the schema where the study population is stored. Most likely a data-type scheme, e.g. [moe\_clean], or a project schema, e.g. [DL-MAA20XX-XX].
- 4. Table Name (required, sql object only) The name of the table/view where the subject population is stored.
- Identity Column (required, sql object or constant) The name of the column that contains the unique identifier for the population. This can be an individual identifier, e.g. [snz\_uid], or some other identifier, e.g. [snz\_xxx\_uid], [mb\_2018], [school\_id].
- Label Identity (required, sql object or constant) Used to label your study population, e.g. "treat" and "control". Use a placeholder like "person" if unneeded.
- Summary Period Start Date (required, sql object or constant) Start date for time of interest for the subject population. If entering constants, the data must be given as "YYYY-MM-DD" e.g. "2020-01-01" as the start of 2020.
- Summary Period End Date (required, sql object or constant) End date for time of interest for the subject population. If entering constants, the data must be given as "YYYY-MM-DD" e.g. "2020-12-31" as the end of 2020.
- 9. Label Summary Period (required, sql object or constant) Used to label your study periods, e.g. "before" and "after". Use a placeholder like the study year "2018" if unneeded.

The assembly tool does not require the input or output to be individual people. Identity\_column could contain geographic or business identifiers. More details on how different types of identities can be used are discussed below under "Non-individual identities".

The start and end date are used to filter all the measures. Only measures that overlap with the summary period for the population will be included in the output. So start date "2020-01-01" and end date "2020-12-31" will exclude measures that sit outside the year 2020 (e.g. income, hospital visits, benefit receipt that ended before 2020, or that started after 2020 will be excluded).

The correct format for dates is YYYY-MM-DD. Other date formats may not result in an error but could lead to incorrect output (for example, depending on the computer's settings DD-MM-YYYY and MM-DD-YYYY dates can be treated as the wrong type).

Define measures fo	or analysis	
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Tool is intended to work with measures that are one row per event	Total income from lages and Salaries. Notes: 1) Saif employment does not appear in this definition. 2) Montantes tos cours lag days in exployment as people working multiple jobs will 3) a placedoidsr identity exists where the encrysted BDO number [sss: Ind sid] is equal to 4) a placedoidsr identity exists where the encrysted BDO number [sss: Ind sid] is equal to	
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Example on RHS is of an employment event. The details of this event include who, when, income paid, employer's identity.

The tool works best when the measures are from tables that are one row per event. It helps that most IDI tables are already one row per event (e.g. hospital discharge, monthly tax summary, school enrolment). But raw table events may not be the events we want to study, and so we make new events from old ones (e.g. hospital discharge event  $\rightarrow$  knee replacement surgery event; OR school enrolment  $\rightarrow$  studying at university event).

I recommend, defining simple measures as Views in IDI\_UserCode. And defining more complex measures as Tables in IDI\_Sandpit. For some very simple events you may be able to use raw data directly.

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	Revised definition	
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deprivation, wrban/rural, and whether a person lives in a household with dependent children.	deprivation, urban/rural, and whether a person lives in a household with dependent children.	
Intended purpose: Identifying the region, whan/rural-mess, and other characteristics of where a person lives at a specific point in time.	Intended purpose: Identifying the region, urban/rural-ness, and other characteristics of where a person lives at a specific point in time.	
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we note that this is a low quality measure. 3) For simplicity this table considers address at a specific date.	we note that this is a low quality measure. 3) for simplicity this table considers address at a specific date.	
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This is also a good opportunity to reuse/revise previous definitions. In the example above, we update some of the parameters of an old definition so it can be reused as a new definition.

Both images show the meta-data header I use for my definitions. One component of this header is a list of parameters that are particular to an analysis (e.g. project prefix, IDI refresh, target date). This makes find & replace an easy means of reusing entire definitions.

When doing this take care to check your inputs for consistency. The user is responsible for ensuring the correct parameters (including choice of IDI refresh) are used everywhere. You will get odd results if you reuse definitions that refer to a different IDI refresh.

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Enter the details of each measure you want in your output. All columns other than File and Notes/description are compulsory.

Configuration of measure file is very similar to the population file. Please refer to notes from configuration of population file.

Columns:

- Database, schema and table names give the SQL location where the table is saved
- Identity is the column that contains individual identifiers
- Measure period start and end date define the period of time of the event (Format: "YYYY-MM-DD", e.g. "2020-12-31")
- Label measure is the labels your output will have (recommended to use underscores instead of spaces)
- The value your output will have is determined by "Measure\_summarised\_by" applied to "Value\_measure"
- Proportional is used to pro rate to the intersection between the summary period and the measure period. It is especially important when working with the duration summary type.

Details on each column of the population file

1. File (optional) – Recommended you record the file name of the SQL script that

creates your measure.

- Database Name (required, sql object only) The name of the SQL database where the measure is stored. Most likely [IDI\_Clean\_XXXX] or [IDI\_Sandpit] or [IDI\_UserCode].
- 3. Schema Name (required, sql object only) The name of the schema where the measure is stored. Most likely a data-type scheme, e.g. [moe\_clean], or a project schema, e.g. [DL-MAA20XX-XX].
- 4. Table Name (required, sql object only) The name of the table/view where the measure is stored.
- Identity Column (required, sql object or constant) The name of the column that contains the unique identifier for the population. This can be an individual identifier, e.g. [snz\_uid], or some other identifier, e.g. [snz\_xxx\_uid], [mb\_2018], [school\_id].
- 6. Measure Period Start Date (required, sql object or constant) Start date for event. If entering constants, the data must be given as "YYYY-MM-DD" e.g. "2020-01-01" as the start of 2020.
- Measure Period End Date (required, sql object or constant) End date for event. If entering constants, the data must be given as "YYYY-MM-DD" e.g. "2020-12-31" as the end of 2020.
- Label Measure (required, sql object or constant) The label assigned to the output measure. It is recommended to avoid special characters and use underscore "\_" instead of space " ".
- 9. Value Measure (required, sql object or constant) The variable name that will be measured. In some cases this will be a placeholder value.
- 10. Measure Summarised By (required, specific list of accepted commands) This column stores the (pre-defined) function that will be used to summarise the measure. The next slide details on what these functions are and what they do.
- 11. Proportional (required, TRUE or FALSE) Proportional is used to pro rate to the intersection between the summary period and the measure period. If a variable is measured with proportional being TRUE, then the measure summary (which is usually duration / number of days) will be calculated for the period falling within the population summary start and end dates.
- 12. Notes / Description (optional) Recommended you record a short description of the variable that is produced.

Value\_measure may take placeholder values. For example, when calculating DURATION a column containing additional values (such as income earned) is not required. Or when counting all records, we count a placeholder "1" as counting a specific column only counts people with a non-missing value for that column. We use "1" as our placeholder.

### Summary types for measures

### Measure\_summarised\_by column

- MIN minimum of the value\_measure column
- MAX maximum of the value\_measure column
- SUM sum of all amounts in the value\_measure column
- MEAN the mean of the value\_measure column
- COUNT number of non-missing values
- DISTINCT number of distinct values
- EXISTS indicator for any records, 1 if COUNT > 0; else 0
- DURATION sum of number of days in events
- HISTOGRAM counts each unique value in the input, generates multiple outputs



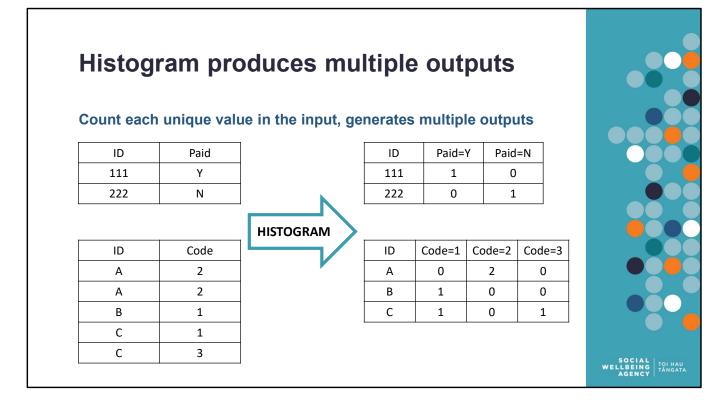
MIN, MAX, SUM, MEAN, DISTINCT and HISTOGRAM apply to Value\_measure column in control file.

DURATION does not use Value\_measure column. We use "1" as a placeholder. COUNT and EXISTS will not count missing values in the Value\_measure column. We use "1", to count all records.

You may need to cast values into numeric type before assembling with the tool. For example, a column might contain codes "00" and "01", but if these are stored as text then they can not be summarised using MIN, MAX, or SUM. They could be summarised using HISTOGRAM, but it is often best to convert/cast these values to numbers when creating the input measures.

Most of these functions translate direct to the equivalent SQL summary function. MEAN can also be created by calculating SUM and COUNT and then dividing SUM by COUNT.

DISTINCT will count missing/NULL columns as a separate type (https://www.sqlservertutorial.net/sql-server-basics/sql-server-select-distinct).

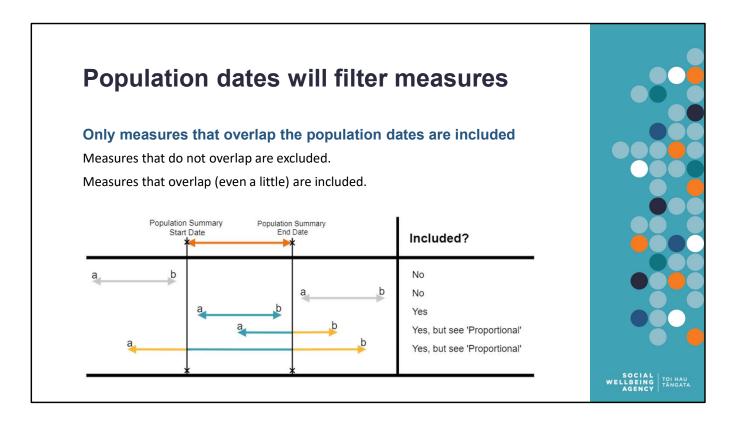


HISTOGRAM can be a good choice for summarising text columns that can not easily be cast into text.

HISTOGRAM can be considered as a "count each type" summary measure. It produces multiple output columns, one for each unique value in the input. This is most useful when the input column contains text codes. Because the assembly tool only outputs numeric values, HISTOGRAM can be used to produce a collection of indicator columns from text. There are helper functions in the supporting R scripts to reverse this and combine multiple columns back into a single one.

HISTOGRAM produces a new column in the output table for each unique value in the value\_measure column. Before using HISTOGRAM, check that there are only as small number of unique values in the column: ~100 values (e.g. TA codes) is fine. More than 1000 unique values will produce errors because SQL has a limit of 1024 columns in a single table.

Researchers who are familiar with R may find the 'collapse\_indicator\_columns' function in dbplyr\_helper\_functions useful for recombining histogram columns back into a single column (where applicable).

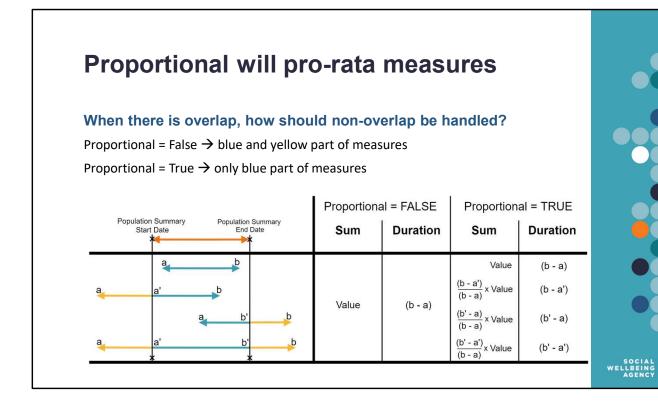


This applies to all summary types. Summary dates for population are compared against dates for measures. Only measures that overlap with the summary period will be included in the output.

Orange gives the summary period for a member of the study population. Grey is excluded, Teal is included, Yellow is included but depends on the Proportional column.

In order from top to bottom in the figure:

- 1. Measures that end before the population summary start date are excluded.
- 2. Measures that start after the population summary end date are excluded.
- 3. Measures that sit entirely within the population summary period (measure starts after population summary starts, and measure ends before population summary ends) are included.
- 4. Measures that start (or end) within the population summary period but end (or start) outside the population summary period are included but are affected by the 'Proportional' column of the control file.
- 5. Measures that start before, and end after, the population summary period are included, but are affected by the 'Proportional' column of the control file.



For DURATION, SUM, and MEAN summary types only. When there is some overlap between a measure and the summary period, Proportional determines how non-overlap is handled. Either we include the entire value/measure or we scale it down (pro-rata) to the amount that overlaps.

Orange gives the summary period for a member of the study population. Teal is included, Yellow depends on the Proportional value. The labels are:

- a the measure start date
- b the measure end date
- a' the population start date used when measure starts <u>before</u> the summary period.
- b' the population end date used when measure ends <u>after</u> the summary period.

The columns to the right shows how the value of SUM or DURATION changes depending on proportional. MEAN changes in that equivalent way to SUM.

When using DURATION you will often want PROPORTIONAL = TRUE.

- PROPORTIONAL = FALSE with DURATION counts all days where the event period has some overlap with the summary period.
- PROPORTIONAL = TRUE with DURATION counts all days in both the event period and the summary period.

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While you can call the assembly tool programmatically from R, the script run\_assembly.R has been created to simplify the most common case: assembling a rectangular dataset from a pair of control files.

Enter the details in the section labelled user controls. For your first run, ensure that development mode is set to TRUE.

You can then run the script

- click the bottom in the top-right of the script window labelled "Source"
- OR press Ctrl + Shift + S

You will start to see console output (info messages for the user) like on the right. Next slide shows the end of the console output.

Note the tool (attempts to) assembles every measure (row in the measures table) for every population (row in the populations table). Hence in the example above, the tool covers 21 measures for 12 populations (it was a panel of 12 quarters).

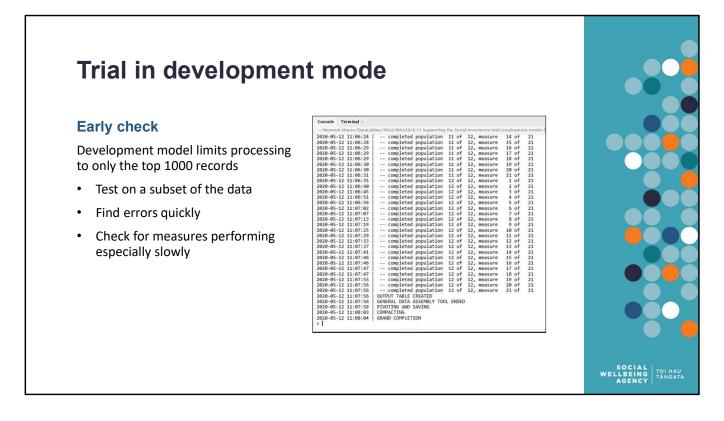
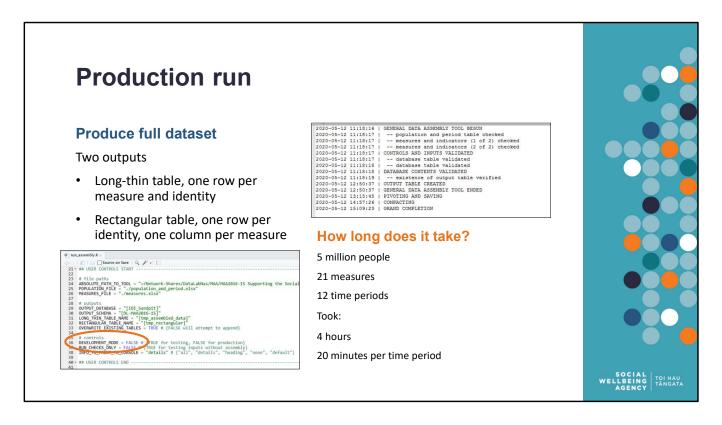


Image gives the end of the console output (info messages for the user) from a run. Once development mode is finishes, it is good practice to inspect the output table in SQL to ensure everything is as expected.

Why development mode? This is a design pattern I use to avoid long wait times during development/testing. Because large datasets can require large amounts of processing time DEVELOPMENT\_MODE = TRUE limits the program to only the top 1000 records from each measure. This helps check that every input dataset performs as expected, but not so large that we wait several hours for a failure. Ideally the tool will finish in <10 minutes in development mode.

A good thing to check from the console log is how long did each measure take to run. Measures that are slow to run may need improvement (more efficient SQL code, convert views to tables, add indexes to tables).

Even if development mode runs without error or concern this does not guarantee producing the full dataset will be error free. We have observed cases when no error occurs with development mode turned on but does occur with development mode turned off, because the record that causes the error is not in the first 1000 records.



Once you have tested using development mode, then you are ready to produce your full dataset.

Setting DEVELOPMENT\_MODE = FALSE means the tool will assemble the entire dataset. This will take longer than development mode.

RHS gives example of a large production run.

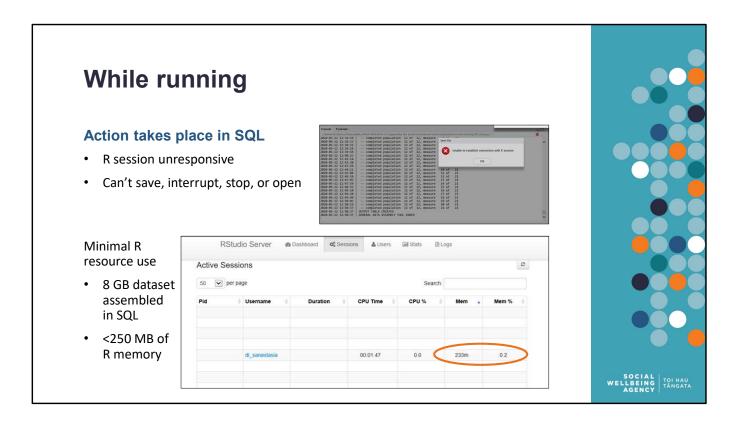
For 5 million NZers, 21 measures, and 12 time periods: Total time taken was 4 hours. This is ~20 minutes per time period (for a single cross-section).

So, an entire panel dataset assembled in a day (but don't sit waiting for it to finish). And, a cross-sectional dataset assembled while you plan the next steps.

run\_assembly.R requires you to specify two output table names.

- A long-thin table directly produced by the tool. It is easiest for the tool to produce one row per identity-and-measure-pair.
- A rectangular table for researchers with one row per identity and one column per measure.

For a worked example see the outline document.



Assemble the entire dataset takes longer than development mode. Perhaps 10 times longer.

Because most of the assembly takes place on the SQL server, the tool puts minimal pressure on R memory or resources.

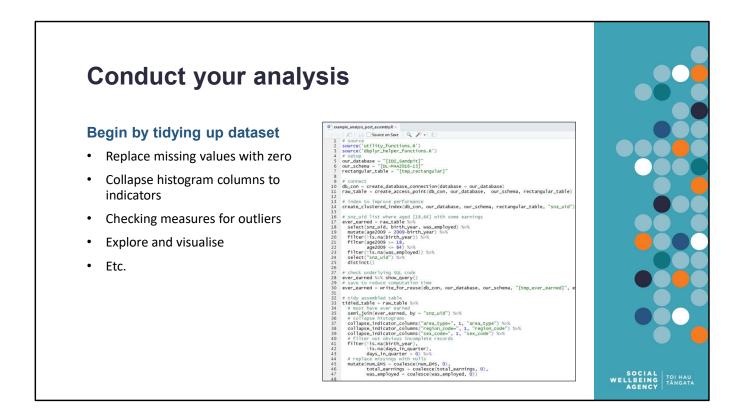
However, while it is running the tool is making consistent use of SQL server resources. So you still need to be considerate of other users – the datalab is a shared resource.

Even though SQL server does most of the work, your R session will still be unresponsive while the tool is running as R waits for SQL.

If you are an R developer, you may want to run two R sessions – one for the tool and the other for development.

- You will need to open the second session before starting the tool.
- You are responsible for closing both R sessions the datalab is a shared resource.

The only way I have found to stop R running when it is waiting for the SQL server to respond is to close the browser (R Studio is accessed in the IDI via a web-browser), reopen it, and log back in. You will then be prompted that R is non-responsive do you want to stop any running process.



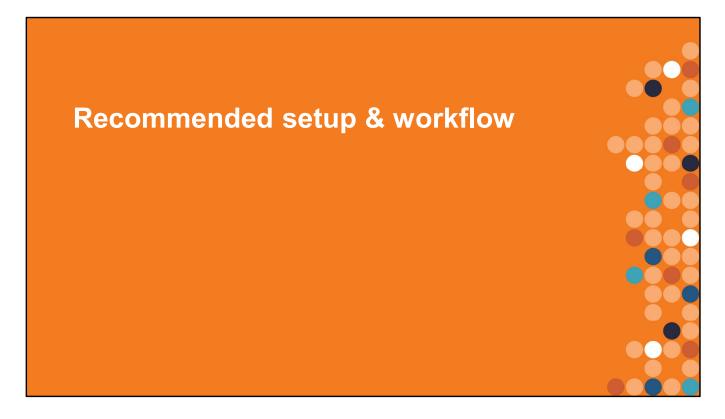
This tool is an <u>assembly</u> tool. Unless you conducted thorough data cleaning before assembly, you will need to conduct it afterwards.

Two most common tasks post-assembly tool are:

- Replace missing values with zeros: If a measure is sum income and a person has no income, then this value will be missing/NULL for them. You will need to decide how to handle missing values. In many cases you will want to replace missing values with zeros.
- Collapse histogram columns: The histogram summary command produces multiple columns, one for every distinct value in the input column. This is necessary because the assembly tool requires numeric outputs as certain stages. These can be collapse back from a series of indicator columns to a single column.

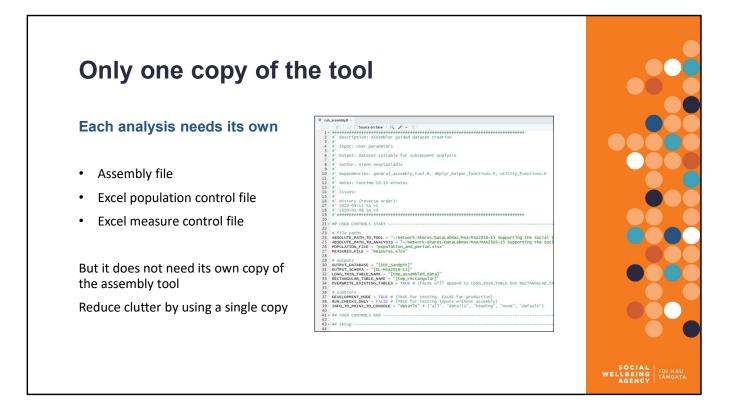
For data exploration and visualisation, I recommend

- 1. Filtering to at most 50,000 rows
- 2. Collecting the data into R
- Visualising it with the explore package (<u>https://cran.r-</u> project.org/web/packages/explore/vignettes/explore mtcars.html)



While you are welcome to use the assembly tool in the way that best suits you (being a good citizen of the shared datalab resources), I have found it to be especially effective as part of a broader workflow pattern.

Consider adopting one or more of the following practices to improve your IDI research.



While it can be tempting to have a copy of the Dataset Assembly Tool for each separate piece of analysis, only one copy of the tool is needed for an entire research project.

run\_assembly.R requires two paths: the location of the tool and the location of the analysis.

- The location of the tool is the folder where general\_assembly\_tool.R and automated\_tests.R are stored
- The location of the analysis is where run\_assembly.R and the Excel control files are stored

When you run the assembly tool, it will first load the tool from the first folder and then carry out the assembly specified in the second folder.

Therefore you can have many Excel control files and many different datasets assembled, all using the same copy of the tool.

### The iterative workflow

#### Iteration is expected

But analysis is smoother when information flow is linear:

 $Raw \rightarrow Prepared \rightarrow Analysed \rightarrow Results$ 

Temptation to churn in place, but adding new raw data onto already analysed data creates confusion

Solution is to rerun assembly process

#### **Exploration workflow**

- 1. Identify the need for a new measure or a change to an existing measure
- 2. Explore IDI data (in SQL)
- 3. Propose and validate new measure
- 4. Define the new measure in SQL as either a table or view
- 5. Add the new measure to control file
- 6. Rerun assembly tool
- 7. Load dataset into analysis
- 8. Resume analysis



The nature of research and development is that it is non-linear: Data preparation and preliminary analysis reveals that other data is needed, so additional data is gathered, further results are produced, and the process iterates. While your process may be iterative and non-linear, the flow of information through your analysis can be linear.

Having a linear flow of information simplifies and clarifies your analysis. The goal is that at each stage all the data is in the same state:

Raw unformed IDI  $\rightarrow$  prepared events  $\rightarrow$  freshly assembled data  $\rightarrow$  cleaned and analysis ready  $\rightarrow$  results

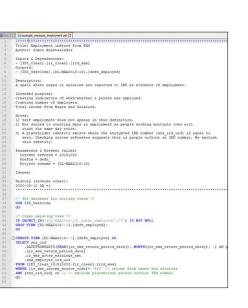
In this way a single dataset is never a mixture of both raw & analysed data. Don't add raw data to analysed data, add raw data to other raw data and analyse it all together.

Because the assembly tool is designed to be rerun, it helps ensure that the initial part of your data process is linear. Designing your data cleaning and analysis to be repeated once new data is added or definitions are changed, helps ensure you maintain this linear process.

### Document and share your definitions

### Stand alone SQL definitions

- Independent of summary tool
- Contain meta-data
- Easy to share & publish
- Easy use & adapt
- Improve IDI for all researchers
- Build a library of definitions



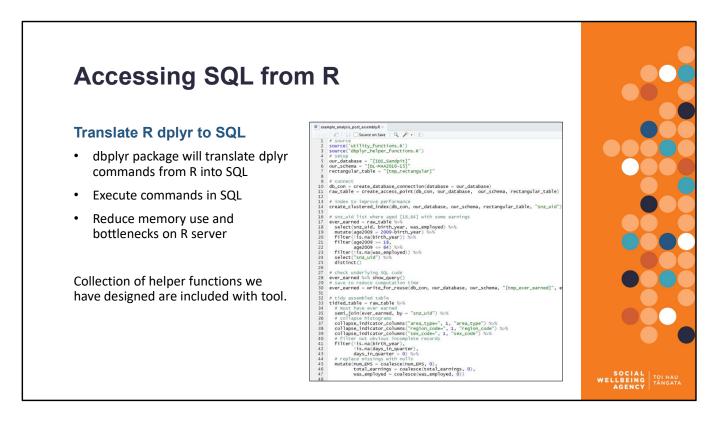


Once you have researched the raw data, constructed a definition for a specific measure, and validated the correctness of the measure it is good practice to document your work. Writing up some documentation will be less effort than having to retrace your steps if you forget, and should take less time than the tool saves you.

Sharing your definitions is part of publishing your code. As more of us do this, we will together build a Library of Definitions. This will be a significant step in the maturity of the IDI. Imagine being able to get health definitions created by health experts, or benefit definitions created by benefit experts. The breadth of the IDI means we need a wide range of subject matter experts – this is a practical way to start.

I propose these definitions are written in SQL. This makes them accessible to both SAS and R users. Even if they prefer R or SAS, most IDI researchers know a little SQL anyway because that is how you view the raw data.

The 'expanded header' is the current documentation approach. Not necessarily an excellent approach, but more important to get started with an approach than to ensure the approach is perfect before starting. As the library of definitions grows we can expect this process will be refined.



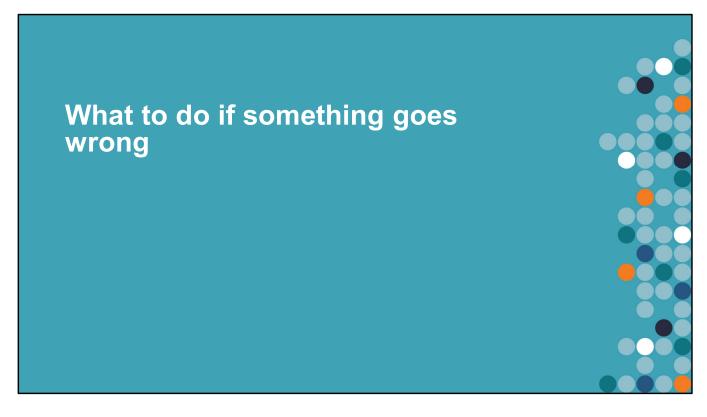
While SQL is well suited to defining populations, events, and measures SAS and R tend to be preferred for programming.

Just as it is possible to connect SAS to SQL, write SAS commands, and have them executed by SQL,

it is also possible to connect R to SQL, write R commands, and have them executed by SQL.

This reduces memory use and bottlenecks on the R server. Uses SQL for its strength – working across large datasets. Collect into R once prepared and tidied to use R for its strength – statistical analysis.

The key R package for doing this is dbplyr. Along with the assembly tool, we also provide a collection of helper functions to support researchers wanting to take this approach. An example using this approach is included in the documentation file (displayed in image).



Nothing is perfect, everything will break eventually. But most errors or problems should be fixable by the end user.

(Low dependence on an external expert, more self-service progress.)

Just because the presentation covers error handling in detail does NOT mean the tool is error prone. This section is detailed because so much effort has gone into ensuring the tool works smoothly for users.

Skip over this section in a short presentation.

### **Program errors and use errors**

#### **Program errors**

- Dataset Assembly Tool does not complete running
- R or SQL gives error message
- Cause is most likely of a technical nature

#### **Use errors**

- Dataset Assembly Tool completes running but output is not as expected
- Unusual output discovered during data exploration
- · Cause is most likely one of misunderstanding

Read the speakers notes in this presentation - they give extra guidance

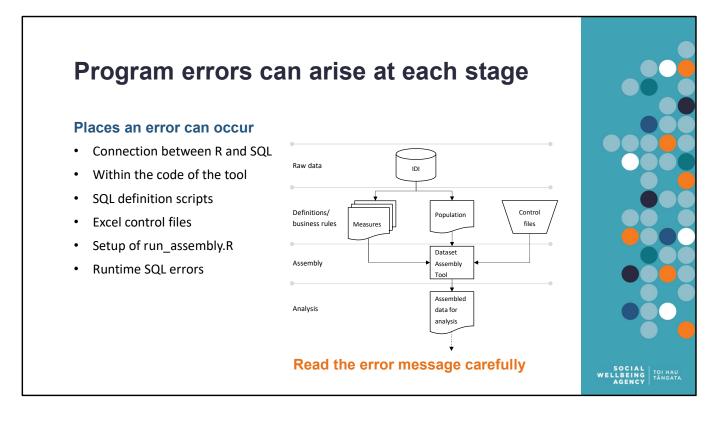
Two broad reasons things can go wrong:

- 1. The computer attempts something it can not do  $\rightarrow$  program errors
- 2. The computer does as instructed but does not give the desired output  $\rightarrow$  use errors

Because the tool reads in definitions <u>created by the user</u> and control files <u>created by the</u> <u>user</u> there is a lot of scope for the user to give the tool invalid or faulty inputs. This section is designed to help users give correct inputs to the tool, and to correct their inputs when things do not work as expected.

For <u>some</u> Use Errors, we have added checks to the tool that will give warnings or errors instead of producing unusual output. This blurs the difference between a Use Error and a Program Error.



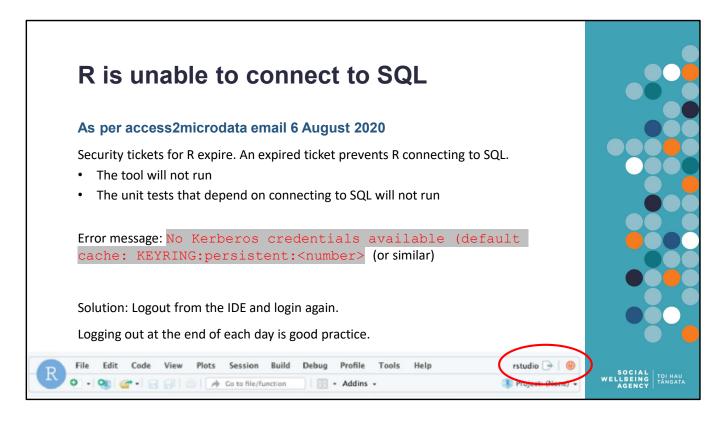


There are a series of steps/stages that are involved in the assembling of data via the tool. Error are possible at each stage.

Program Errors at each of the different stages look different and have different solutions.

- Program Errors at all of the stages can be identified without R / programming knowledge.
- Program Errors at most of the stages can be fixed without R / programming knowledge.

Always read the error message, the tool has been designed to provide instructive error messages and user feedback.



We most often encountered this error message when returning to R after leaving it logged in overnight.

This has been consistently resolved for us by (1) ending the current session (red 1/0 circle) and (2) logging out from RStudio (square & arrow symbol).

Both of these are good practice for finishing with & tidying up from an R session (releases R memory and restarts you with a clean environment).

Text from Access2microdata email 6 August 2020:

### If you experience issues with ODBC:

- If you get the error "No Kerberos credentials available (default cache: KEYRING:persistent:<number>)", then logout from the Rstudio Pro IDE and login again, as it is a security requirement that Kerberos tickets expire. Logging out at the end of the day is good practice, anyway.
- 2. Any errors referring to the ODBC driver version 11 not found, use: Driver = "ODBC Driver 17 for SQL Server".
- 3. Any other ODBC errors: check if you have a personal copy of an old odbc CRAN package. If so move it away. Then the system version of the odbc CRAN package (/usr/share/R/library/odbc) will be used, which has been proven to work.

### No error but unresponsive

#### Valid R, valid SQL, but server unresponsive

- The IDI is a shared environment
- The IDI involves networked resources

Hence other programs running on the server, or in the network, can interfere with the Dataset Assembly Tool.

- These problems usually resolve, once the other program finishes running
- Where problems persist or repeat, it may help to inform the Integrated Data team so they can look into the cause



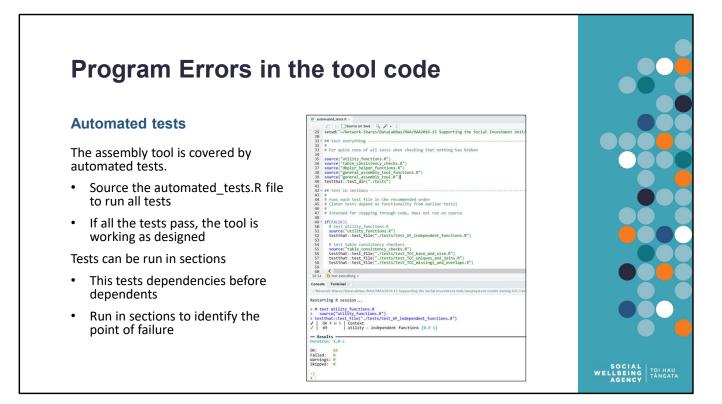
Poor performance / non-responsiveness is unlikely to be an error, and more likely to be high traffic or large programs.

In August 2020, the most common problem of this nature was R becomes nonresponsive because it is waiting for a response from SQL. If, while waiting for R, we went into SQL and requested a list of all Sandpit tables (a task that usually takes 1-2 seconds) we would instead receive an error message. This highlights that the problem is not with R or with the tool because SQL also errors.

Good checks to run:

- 1. Check you can use R without the tool
- 2. Check you can carry out equivalent work in SQL
- 3. Ask colleagues if they are experiencing the same or similar problems

When problems persist or repeat, you could inform the Integrated Data team in case they are unaware of the problem. When doing so we recommend providing details to assist their staff identify the cause (which may be you). This will not resolve the problem immediately, but it helps to improve the performance and robustness of the IDI environment – something that both Stats NZ and IDI researchers want.



Take a look at the image:

- The top third, lines 29-40, tests everything (but in no specific order). This code runs when the file is sourced.
- The middle third, lines 42-onwards, tests each section of the code in the order of dependency (later code will fail if earlier code does not pass). Run this code manually, line by line, is recommended if testing everything produces errors. This can be done by placing the cursor on the line you want to run and pressing Ctrl + Enter.

Note that line 49 is an if(FALSE) statement that prevents testing in sections running when testing everything. Do not run this line if you are trying to test in sections.

• The bottom third, the console, is an example of testing in sections. Here we have run lines 50, 51, and 52 to test the independent functions found in utility\_functions.R

# Population and measure definitions

### These are your responsibility

Tool assembles <u>your</u> tables/views

Run SQL scripts before assembly tool

Common things to check: three core parameters

• Prefix

• Project schema

• IDI refresh

Helpful tricks

- Simple definitions as Views in UserCode
- Complex definitions as Tables in Sandpit
- Index large Sandpit tables
- Run in Development Mode first



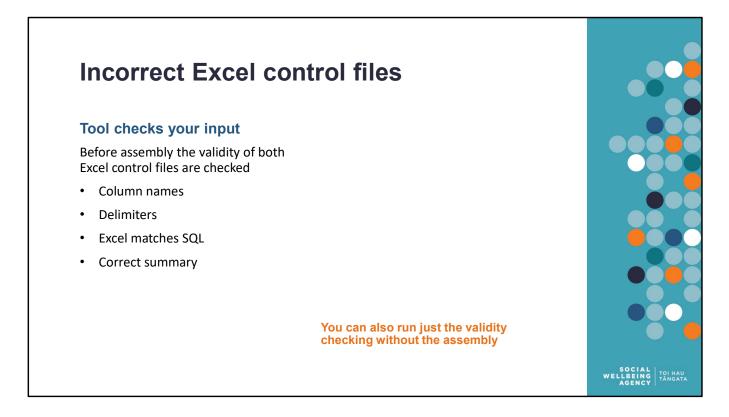
Population and measure definitions are defined in SQL. They are prepared and run before being included in the tool assembly.

- Make sure your SQL definition files run
- Make sure you run all your SQL definitions before running the assembly tool
- Check your definitions for
  - Correct prefix, schema, refresh
  - You have not accidentally restricted yourself to the "TOP 1000" rows

Just because a definition runs does not mean it will perform well.

- Simple definitions should be created as views reduce memory space and make use of existing indexes
- Complex definitions should be created as tables with indexes

If the error is in your SQL definition (population or measure) script, then it is not an assembly tool related error.



Before any dataset assembly happens the tool first checks that all your control file inputs are correct. Both control files are checked.

Four main checks are run:

- 1. The control files have all the required columns
- 2. All the entries in the required columns are correctly delimited
- 3. The SQL tables and columns specified in the control file exist in SQL
- 4. Summarising of variables is correct (e.g. you can't sum text)

It will also give warnings if your summary input is inconsistent (e.g. there is no need to set Proportional = TRUE when counting).

You can run just the validity checking without the assembly. This helps with checking whether your Excel inputs are correct without having to wait for the assembly to run if they are.

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File		[personal detail]	Identity_colum [snz_uid]	Start_date "2008-01-01"	<ul> <li>Measure_period_end_date</li> <li>2010-12-31*</li> </ul>	"birth month"	<ul> <li>Value_measure [snz birth month nbr]</li> </ul>	<ul> <li>Measure_summarised_by</li> <li>MAX</li> </ul>	* Proportional FALSE	NOTES/DESCRIPTION     Month of birth date	
[IDI_Clean_20191020]		[personal_detail]	[snz_uid]	2008-01-01"	2010-12-31*	"birth_year"	[snz_birth_year_nbr]	MAX	FALSE	Year of birth date	
[IDI_Clean_20191020]	[data]	[personal_detail]	[snz_uid]	"2005 "1-01"	*2010-12-31*	"eth_asian"	[snz_ethnicity_grp4_nbr]	MAX	FALSE	Indicator for Asian ethnicity	
[IDI_Clean_20191020]		[personal_detail]	[snz_uid]	"2008-01-01"	*2010-12-31*	"eth_european"	[snz_ethnicity_grp1_nbr]	MAX	FALSE	Indicator for European ethnicite	
[IDI_Clean_20191020] [IDI_Clean_20191020]		[personal_detail] [personal_detail]	[snz_uid] [snz_uid]	"2008-01-01" "2008-01-01"	"2010-12-31" "2010-12-31"	"eth_maori" "eth_MELAA"	[snz_ethnicity_grp2_nbr] [snz_ethnicity_grp5_nbr]	MAX	FALSE	Indicator for Maori ethnicity Indicator for MELAA ethnicity	
[IDI_Clean_20191020] [IDI_Clean_20191020]		[personal_detail]	[snz_uid]	"2008-01-01"	*2010-12-31*	[eth_other]	[snz_ethnicity_grp5_nbr]	MAX	FALSE	Indicator for Other ethnicity	
[IDI_Clean_20191020]	[data]	[personal_detail]	[snz_uid]	"2008-01-01"	"2010-12-31"	"eth_pacific"	[snz_ethnicity_grp3_nb]	MAX	FALSE	Indicator for Pacific ethnicity	
[IDI_Clean_20191020]		[personal_detail]	[snz_uid]	"2008-01-01"	*2010-12-31*	"sex_code"	[snz_sex_code]	HISTOGRAM	FALSE	Sex, 1 = male, 2 = female	
[IDI_Clean_20191020]		[personal_detail]	[snz_uid]	"2008-01-01"	"2010-12-31"	"days_in_quarter"		DURATION	TRUE	Number of days in quarter	
	[DL-MAA2016-15]	[defn_employed]	[snz_uid] [snz_uid]	[period_start_date] [period_start_date]	[ir_ems_return_period_date] [ir_ems_return_period_date]	"total_earnings" "was employed"	[ir_ems_gross_earnings_amt]	SUM EXISTS	FALSE	total W&S earnings from quarte indicator for whether or not pe	
		[defn_address_descriptors]		"2010-12-31"	"2010-12-31"	"region_code"	[ant_region_code]	HISTOGRAM	FALSE	Region of address at end of 201	
		[defn_address_descriptors]		"2010-12-31"	*2010-12-31*	"SA2_code"	[SA22018_V1_00]	MAX	FALSE	Statistical area 2 code	
address_c [IDI_Sandpit]	[DL-MAA2016-15]	[defn_address_descriptors]	[snz_uid]	"2010-12-31"	"2010-12-31"	"area_type"	[IUR2018_V1_00_NAME]	SUM	FALSE	Type of area of address at end e	
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Here is an example where the control file has an incorrect column name (but this would also happen if a column was missing).

The error message tells you that the column is missing (read the error messages).

Note that case (upper/lower) does not matter. And that the File and Notes/description columns are optional.

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4 [IDI_Clear_20191020 5 [IDI_Clear_20191020		personal_detail] personal_detail]	[snz_uid] [snz_uid]	"2008-01-01" "2008-01-01"	"2010-12-31" "2010-12-31"	"eth_asian" "eth_european"	[snz_ethnicity_grp4_nbr] [snz_ethnicity_grp1_nbr]	MAX	FALSE	Indicator for Asian ethnicity Indicator for European ethnicity	
6 [IDI_Clear_20191020		personal_detail]	[snz_uid]	"2008-01-01"	"2010-12-31"	"eth maori"	[snz_ethnicity_grp1_nbr]	MAX	FALSE	Indicator for European ethnicity	
7 [IDI_Clean_20191020		pers nal_detail]	[snz_uid]	"2008-01-01"	*2010-12-31*	"eth_MELAA"	[snz_ethnicity_grp5_nbr]	MAX	FALSE	Indicator for MELAA ethnicity	
8 [IDI_Clean_2011020		ersonal detail]	[snz_uid]	"2008-01-01"	*2010-12-31*	[eth_other]	[snz_ethnicity_grp6_nbr]	MAX	FALSE	Indicator for Other ethnicity	
9 [IDI_Clean_20191020 10 [IDI_Clean_20191020		personal_detail]	[snz_uid]	"2008-01-01" "2008-01-01"	"2010-12-31" "2010-12-31"	"eth_pacific"	[snz_ethnicity_grp3_nb]	MAX	FALSE	Indicator for Pacific ethnicity Sex, 1 = male, 2 = female	
10 [IDI_Clean_20191020 11 [IDI_Clean_20191020		personal_detail]	[snz_uid] [siz_uid]	"2008-01-01"	"2010-12-31"	"sex_code" "days_in_quarter"	[snz_sex_code]	DURATION	TRUE	Sex, 1 = male, 2 = female Number of days in guarter	
2 measure_[IDI_UserCode]	[DL-MAA2015-15]		[sn_uid]	[period start date]	[ir ems return period date]	"total earnings"	[ir ems gross earnings amt]	SUM	FALSE	total W&S earnings from guarte	
3 measure_[IDI_UserCode]	[DL-MAA20.6-15] [		[sn_uid]	[period_start_date]	[ir_ems_return_period_date]	"was_employed"	"1"	EXISTS	FALSE	indicator for whether or not pe	
14 address_c [IDI_Sandpit]	[DL-MAA2010 15] [	defn_address_descriptors]	[s z_uid]	"2010-12-31"	"2010-12-31"	"region_code"	[ant_region_code]	HISTOGRAM	FALSE	Region of address at end of 201	
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After fixing the previous error and rerunning, we get a new error message.

Here is an example where the delimiters are incorrect. In one case there are no delimiters, and in the other we have used " instead of [ ].

The error message tells you that delimiters are missing, how many, and where they are located.

Reminder:

- If the cell contents are SQL tables and columns then delimited []
- If the cell contents are the value to use then delimit in "
- Measure\_summarised\_by and Proportional columns are not delimited

Note that double quotes is different from single quotes.

And straight double quotes are different from curly double quotes – if copy-and-paste from word check carefully that your quotes are straight not curly.

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1 File * Database name * 5	Schema name Table name	· Identity column - Start date	<ul> <li>Measure period end</li> </ul>		Value_measure	* Measure summaris	ed by Proportional *	NOTES/DESCRIPTION	
2 [IDI Clean 20191020] [		[snz uid] "2008-01-01"	*2010-12-31*	"birth month"	[snz birth month nbr]	MAX	FALSE	Month of birth date	
3 [IDI_Clean_20191020] [		[snz_uid] "2008-01-01"	"2010-12-31"	"birth_year"	[snz_birth_year_nbr]	MAX	FALSE	Year of birth date	
4 [IDI Clean 20191020] [		[snz uid] "2008-01-01"	*2010-12-31*	"eth asian"	and the second second	MAX	FALSE	Indicator for Asian ethnicity	
5 [IDI_Clean_20191020] [		[snz_uid] "2008-01-01"	*2010-12-31*	"eth uropean"	[snz_ethnicity_grp1_nbr]	MAX	FALSE	Indicator for European ethnicity	
6 [IDI Clean 20191020]		[snz_uid] "2008-01-01"	"2010-12-31"	eth_maori"	[snz_ethnicity_grp2_nbr]	MA	FALSE	Indicator for Maori ethnicity	
7 [IDI Clean 20191020] [		[snz uid] "2008-01-01"	*2010-12-31*	"eth MELAA"	[snz ethnicity grp5 nbr]	MAX	FALSE	Indicator for MELAA ethnicity	
8 [IDI_Clean_20191020] [		[snz_uid] "2008-01-01"	*2010-12-31*	[eth_other]	[snz_ethnicity_grp6_nbr]	MAX	FALSE	Indicator for Other ethnicity	
9 [IDI_Clean_20191020] [		[snz_uid] "2008-01-01"	*2010-12-31*	"eth_pacific"	[snz_ethnicity_grp3_nb]	MAX	FALSE	Indicator for Pacific ethnicity	
10 [IDI_Clean_20191020]	Idata) Inersonal detaill	[snz uid] "2008-01-01"	*2010-12-31*	"sex code"	[sna sex code]	HIP OGRAM	FALSE	Sex, 1 = male, 2 = female	
11 [IDI_Clean_20191020]	Console Terminal ×							imber of days in quarter	
12 measure_[IDI_UserCode]							-0	tal W&S earnings from quarte	
13 measure_[IDI_UserCode]	~/Network-Shares/DataLab!	Nas/MAA/MAA2016-15 Supportin	g the Social Investment Unit	t/employment trends	s during GFC/rprogs/ 🛋			dicator for whether or not pe	
14 address_c [IDI_Sandpit]	> source( 'w/Network-St	hares/DataLabNas/MAA/MAA2	016-15 Supporting the	e Social Investe	ment Unit/employment	nt trends during	g GEC/rnrogs/	gion of address at end of 201	
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		ty_grp3_nb] missing from	table [personal_deta:	i1]					
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After fixing the previous error and rerunning, we get a new error message.

Here is an example where the tool can not find the columns specified in SQL.

- For [eth\_other] this is because there is no column [eth\_other] this should be a label "eth\_other" and is incorrectly delimited
- For [snz\_ethnicity\_grp3\_nb] this is a typo, the column name has been incorrectly entered (it should end \_nbr] not \_nb])

The error message gives you the value of the cell that can not be found.

If you program SQL with square brackets (as SQL defaults to when you ask to view the top 1000 rows) then the best way I have found of populating these cells is:

- double-click on a variable in SQL
- Сору
- and paste into Excel

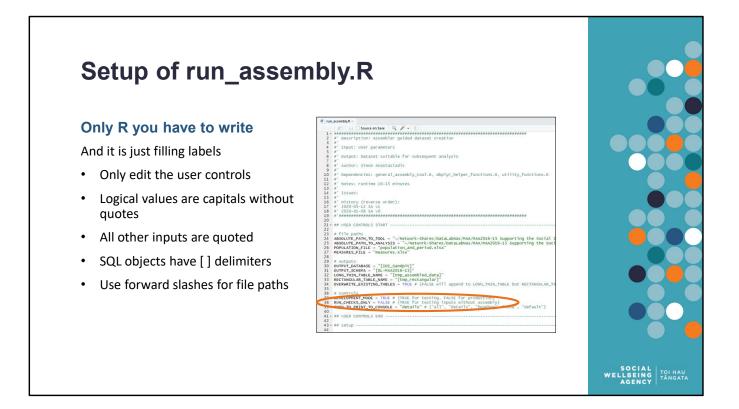
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File • Database_name •	Schema_name -	Table_name •	Identity_column	Start_date	Measure_period_end_date	Label_measure	Value_measure	* Measure_summarised_b	Proportional	<ul> <li>NOTES/DESCRIPTION</li> </ul>	
[IDI_Clean_20191020]			[snz_uid]	"2008-01-01"	*2010-12-31*	"birth_month"	[snz_birth_month_nbr]	MAX	FALSE	Month of birth date	
[IDI_Clean_20191020]			[snz_uid]	"2008-01-01"	"2010-12-31"	"birth_year"	[snz_birth_year_nbr]	MAX	FALSE	Year of birth date	
[IDI_Clean_20191020]			[snz_uid]	"2008-01-01"	"2010-12-31"	"eth_asian"	[snz_ethnicity_grp4_nbr]	MAX	FALSE	Indicator for Asian ethnicity	
[IDI_Clean_20191020]			[snz_uid]	"2008-01-01"	*2010-12-31*	"eth_european"	[snz_ethnicity_grp1_nbr]	MAX	FALSE	Indicator for European ethnicity	
[IDI_Clean_20191020]			[snz_uid]	"2008-01-01"	"2010-12-31"	"eth_maori"	[snz_ethnicity_grp2_nbr]	MAX	FALSE	Indicator for Maori ethnicity	
[IDI_Clean_20191020]			[snz_uid]	"2008-01-01" "2008-01-01"	"2010-12-31" "2010-12-31"	"eth_MELAA" [eth_other]	[snz_ethnicity_grp5_nbr]	MAX	FALSE	Indicator for MELAA ethnicity Indicator for Other ethnicity	
[IDI_Clean_20191020] [IDI_Clean_20191020]			[snz_uid] [snz_uid]	"2008-01-01"	"2010-12-31"	"eth pacific"	[snz_ethnicity_grp6_nbr] [snz_ethnicity_grp3_nb]	MAX	FALSE	Indicator for Dacific ethnicity	
[IDI_Clean_20191020] [IDI_Clean_20191020]			[snz_uid]	"2008-01-01"	"2010-12-31"	"sex code"	[snz_sex_code]	HISTOGRAM	FALSE	Sex, 1 = male, 2 = female	
[IDI_Clean_20191020]			[snz_uid]	"2008-01-01"	"2010-12-31"	"days_in_quarter"		DURATION	TRUE	Number of days in guarter	
			[snz_uid]	[period start date]	[ir ems return period date]	"total earnings"	[ir_ems_gross_earnings_amt]	SUM	FALSE	total W&S earnings from guarte	
			[snz_uid]	[period_start_date]	[ir_ems_return_period_date]	"was_employed"		EXISTS	FALSE	indicator for whether or not pe	
		[defn_address_descriptors]		"2010-12-31"	"2010-12-31"	"region_code"	fant region code]	HISTOGRAM	FALSE	Region of address at end of 201	
		[defn address descriptors]		"2010-12-31"	"2010-12-31"	"SA2 code"	[SA22018_V1_00]	MAX	FALSE	Statistical area 2 code	
		[defn address descriptors]		"2010-12-31"	"2010-12-31"	"area type"	[IUR2018 V1 00 NAME]	SUM	ALSE	Type of area of address at end e	
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>	1										AGENCY

After fixing the previous error and rerunning, we get a new error message.

Here is an example where the contents of the column containing the value\_measure are not compatible with the measure\_summarised\_by that has been chosen. The error message tells you what column, what command, and why.

Many codes in the IDI are stored as text rather than as numeric. This is the most common cause I have encountered. Two solutions:

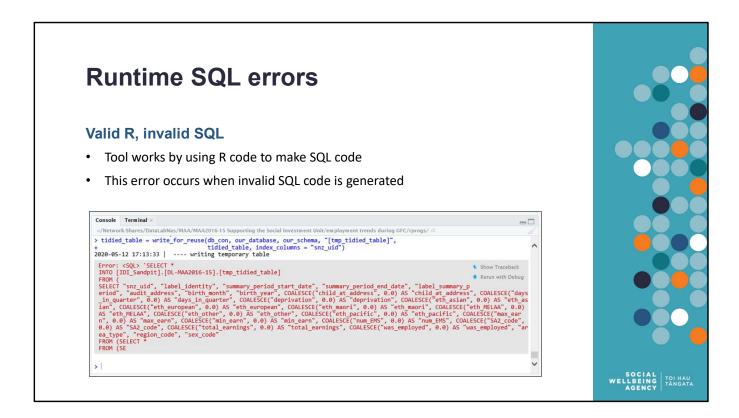
- Cast the variable to numeric in SQL: CAST(COL AS TYPE) AS new\_name
- Use the histogram summary instead



This is the only piece of R code you have to edit, and all you are doing is entering your inputs.

- Only edit within the section labelled user controls (lines 21-40)
- Logical values are in capitals, no quotes
- Everything else is quoted
- SQL objects (database, schema, table names) have [] delimiters without their quotes (e.g. "[output\_database]")

Careful with slashes in file paths. Windows uses backward \ slashes, but R uses forward / slashes. This will cause errors if copy-and-paste from windows without editing.



Sometimes the combination of the input definitions (population and measure) and the SQL code generated by the assembly tool produce errors.

Note how the message starts with "Error: <SQL>" this is a clear sign of a Runtime SQL error. Also observe that the error happened while writing a table. Because writing a table is an SQL activity, this also tells us the error occurred in SQL.

Possibilities include:

- Attempting to SUM a column that is not numeric → The SQL definition and control file both seem correct, but when combining them (text column from SQL definition, SUM instruction from control file) you get an error (the tool now checks for this).
- Too many variables → SQL has a maximum of 1024 columns, so if you create lots of measures (or use HISTOGRAM a lot) you can cause this error. You will need to reduce the number of measures or find a better way to express you current measures.
- Numeric overflow → SQL has a maximum sized integer number it can consider, so if you create a very large value (such as total income ever recorded for all people in the South Island) you can cause this error. You will need to scale down your value (calculate in \$millions) or find another way to measure it.

# "SQL tmp scripts" folder

#### **Review in SQL**

R tells you where the error occurs, but not why

- Open "SQL tmp scripts" folder
- Run the latest code file in SQL Server Management Studio
- Determine from the error message where the problem is

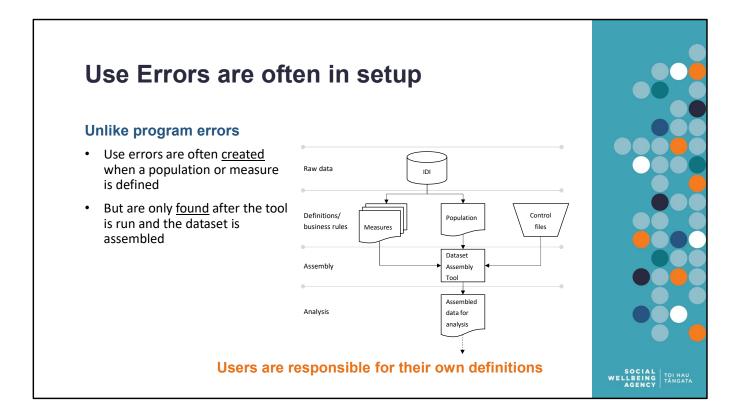
Name	Date modified	Type	Size
documentation	12/05/2020 10:10	File folder	
] SQL tmp scripts	12/05/2020 12:50	File folder	
tests	12/05/2020 10:04	File folder	
automated_tests.R	12/05/2020 9:57 AM	R.File	4 K3
dbplyr_helper_functions.R	12/05/2020 9:56 AM	R File	27 K3
general_assembly_tool.R	4/03/2020 4:45 PM	R File	20 Ki
general_assembly_tool_functions.R	4/03/2020 4:43 PM	R File	5 K3
C measures.xlsx	12/05/2020 10:53	Microsoft Excel W	11 KE
measures_w_errors.xlsx	12/05/2020 1:02 PM	Microsoft Excel W	11 KE
population_and_period.xlsx	12/05/2020 10:09	Microsoft Excel W	9 KE
run_assembly.R	12/05/2020 11:18	R File	4 KE
table_consistency_checks.R	8/05/2020 4:22 PM	R File	12 KE
utility_functions.R	8/05/2020 4:12 PM	R File	7 KI
Name ^	Date modified	Туре	Size
2020-05-12 105259-429 delete_table.sql	12/05/2020 10:52	Microsoft SQL Ser	1.KB
2020-05-12 105259-770 create_table.sql	12/05/2020 10:52	Microsoft SQL Ser	T KB
2020-05-12 105300-228 append_table.sql	12/05/2020 10:53	Microsoft SQL Ser	1 KB
2020-05-12 105305-863 append_table.sql	12/05/2020 10:53	Microsoft SQL Ser	1 KB
2020-05-12 105310-617 append_table.sql	12/05/2020 10:53	Microsoft SQL Ser	1 KB
2020-05-12 105316-091 append_table.sql	12/05/2020 10:53	Microsoft SQL Ser	1 KB
2020-05-12 105321-856 append_table.sql	12/05/2020 10:53	Microsoft SQL Ser	1 KB
2020-05-12 105326-243 append_table.sql	12/05/2020 10:53	Microsoft SQL Ser	1 KB
2020-05-12 105330-303 append_table.sql	12/05/2020 10:53	Microsoft SQL Ser	1 KB
2020-05-12 105334-944 append_table.sql	12/05/2020 10:53	Microsoft SQL Ser	3 KB
2020-05-12 105340-423 append_table.sql	12/05/2020 10:53	Microsoft SQL Ser	1 KB
2020-05-12 105345-542 append_table.sql	12/05/2020 10:53	Microsoft SQL Ser	2 KB
2020-05-12 105350-125 append_table.sql	12/05/2020 10:53	Microsoft SQL Ser	1 KB



When this happens find the "SQL tmp scripts" folder. Every (non-trivial) SQL command executed by the tool gets saved in this folder. Sort the files and open the latest one in SQL Server Management Studio. This should be the code that caused the error. Test whether the code can be run directly from SQL.

- If it runs fine, then there may not be an error just a random hiccup try running the assembly tool again
- If it takes a very long time to run, then it probably takes too long to run for R and the connection is timing out
- If it creates errors, then further inspection is necessary to determine which part of the query is causing the error

You can delete the "SQL tmp scripts" folder without risk – it will recreate itself. We recommend that you delete the folder regularly as it is only there to support debugging / logging and can accumulate thousands of little files which have little value.



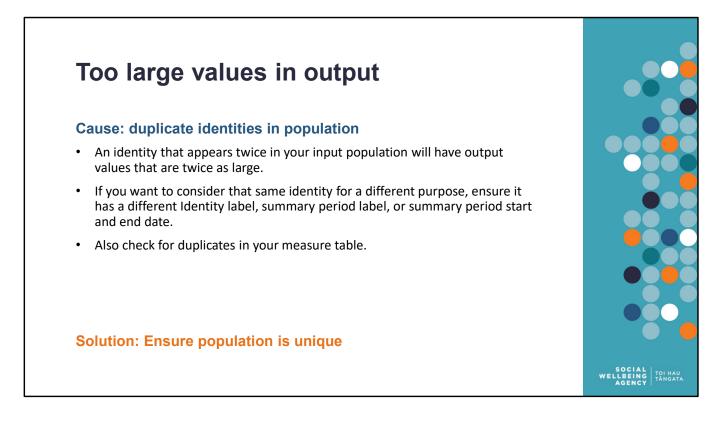
Once output is assembled, it is important to assess the prepared table before it is used in analysis. This often happens as part of tidying up the dataset for analysis (handling missing values, reclassifying variables, removing identities with errors, etc.).

It is not always possible to spot Use Errors from the reduced output produced when the tool is run in DEVELOPMENT MODE. You will often have to fully assemble the dataset before you can check for use error.

Where the assembled dataset contains output that is different from expectations, then further investigation will likely be required.

- Auditing (manually reconstructing) the values for a single identity an effective way to identify user error
- When high standards of quality are critical, you may want to audit the assembled dataset even if there is no sign of use error

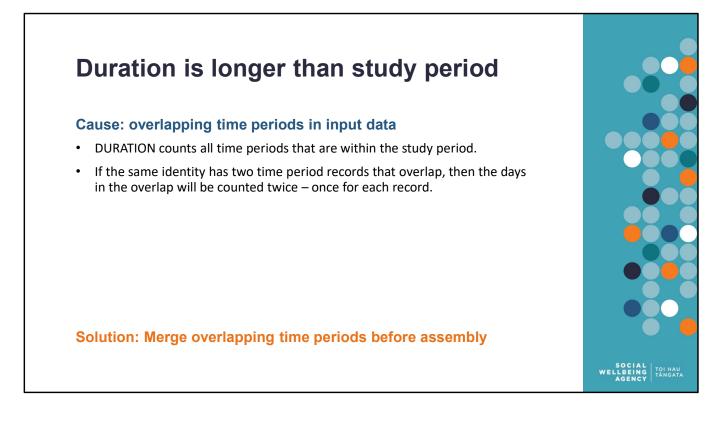
In the following slides, we discuss several types of use error that we are already aware of. More examples may be added later as the number of people using the tool increases.



Because the population table and the measure tables are joined using "INNER JOIN" duplicates in either the population or measure table will get picked up more than once in your output.

Example: Research considers mothers who have a baby born in 2017 or 2018. Initial input population is all mothers in MOH maternity records with year given birth 2017 or 2018. What about mothers who gave birth two (or more) times?

- 1. Do nothing  $\rightarrow$  Duplicate output for these mothers
- 2. De-duplicate the population  $\rightarrow$  One output row for each mother, but mothers who gave birth twice could have twice as many events as mothers who only gave birth once
- Set the summary period start and end date based on the date of the birth → Mothers who gave birth twice appear twice in output dataset, once for each set of dates



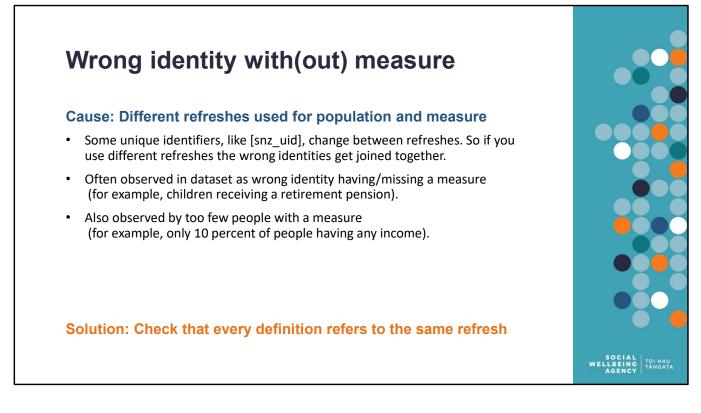
DURATION sums up the individual durations of all time periods that are within the study period. It does not check for whether any of the individual durations overlap. This can produce unexpected output such as 400+ days of employment within a single year.

The solution is to merge your overlapping time periods before assembly.

You can find exemplar code for merging spells in the definitions library SWA has published alongside the assembly tool:

https://github.com/nz-social-wellbeing-agency/definitions\_library

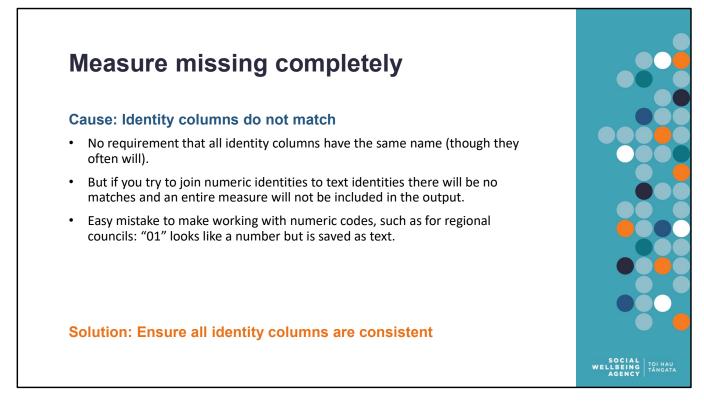
Good existing definitions to check are employment spells, education spells, or benefit spells. In these SQL definitions we often refer to the merging of overlapping time periods as "Condensing spells".



Research should be done using a consistent IDI refresh. When some definitions come from one refresh, and other refreshes some from another, then identity codes can change between refreshes.

The best way we currently have to check this is searching all SQL definition files for "IDI\_Clean\_". Update and rerun any files that are using the wrong refresh.

One supporting tool we hope to see in the future will reduce the risk of this error: A tool that finds the refresh and project schema for every SQL definition. This would make checking for consistency between files much easier.

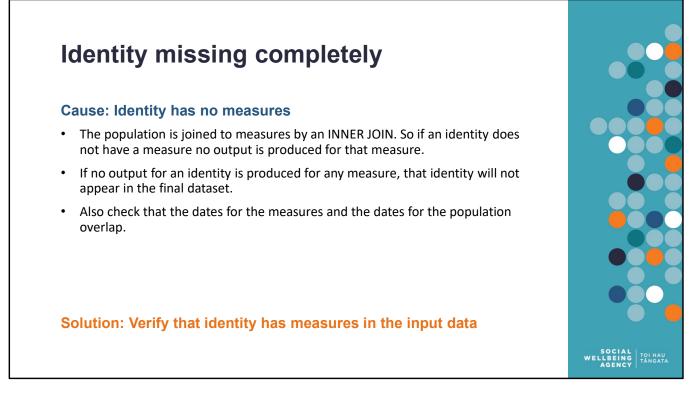


The names of identity columns can be different but the format must be the same

- It is critical to ensure that all the identity columns are in same data format.
- Mismatch of datatypes might not always throw error and can lead to incorrect joins.

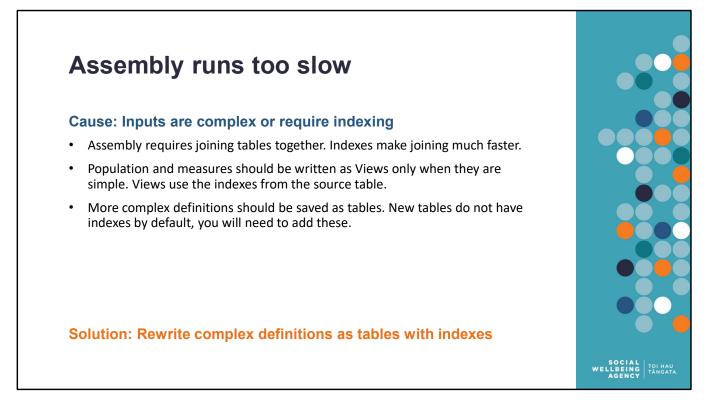
In some special cases SQL will join numeric and text data. If the identities are smaller numbers (for example regional council codes, which are 2 digits), some codes may join: the text "10" may join to the number 10. But other codes will not join: the text "01" will not join to the number 1.

By default [snz\_uid] values are all numeric so this is unlikely to be an problem if using [snz\_uid]



If an identity does not have any measures it will not appear in the final output. This is most likely to occur when working with identities that are not linked to the IDI spine, are not in the residential population, or are 'rare' in some way.

If you want to ensure that an identity is included in the output dataset the easiest was is to include the study population in the measure table. Create a new measure that checks if identities in the population table EXISTS in the population table.



The speed of the assembly tool depends on how fast it is to access and combine the population and measure definitions. Definitions that are slow to access will result in slow performance for the assembly tool.

When running the tool in DEVELOPMENT MODE check for measures that perform slowly. Consider recoding these measures to be more efficient. Adding (clustered) indexes on the identity column improves the assembly speed by a lot.

If you are not sure what definitions are simple and can be Views, and what definitions are too complex and should be tables, take a look at some existing measure definitions. These should also contain code for adding indexes to your tables.

Produ	icing	panel d	lata is (	aitticuit		
		1				
Solution:	Setup vo	ur populatio	on control f	file like this:		
_	Identity_column La			ummary_period_end_date		
[defn_res_father]	[snz_uid]		019-01-01"	"2019-12-31"	"2019"	
[defn_res_father]	[snz_uid]	"father" "2	020-01-01"	"2020-12-31"	"2020"	
[defn_res_father]	[snz_uid]	"father" "2	019-01-01"	"2019-12-31"	"2019"	
[defn_res_father]	[snz_uid]	"father" "2	020-01-01"	"2020-12-31"	"2020"	
[defn_res_mother]	[snz_uid]	"mother" "2	019-01-01"	"2019-12-31"	"2019"	
[defn_res_mother]	[snz_uid]	"mother" "2	020-01-01"	"2020-12-31"	"2020"	
[defn_res_mother]	[snz_uid]	"mother" "2	019-01-01"	"2019-12-31"	"2019"	
[defn_res_mother]	[snz_uid]	"mother" "2	020-01-01"	"2020-12-31"	"2020"	
To motion	and automa					
To get pa	nel outpu	t like this:				
Identity_column	Label_identity	Label_summary	Total_income			
1111	father	2019	3839			
1111	father	2020	9354			
2222	father	2019	4449			
2222	father	2020	4492			
	mother	2019	5204			
3333						
		2020	5032			
3333 3333 4444	mother	2020 2019	5032 3968			SOCIAL

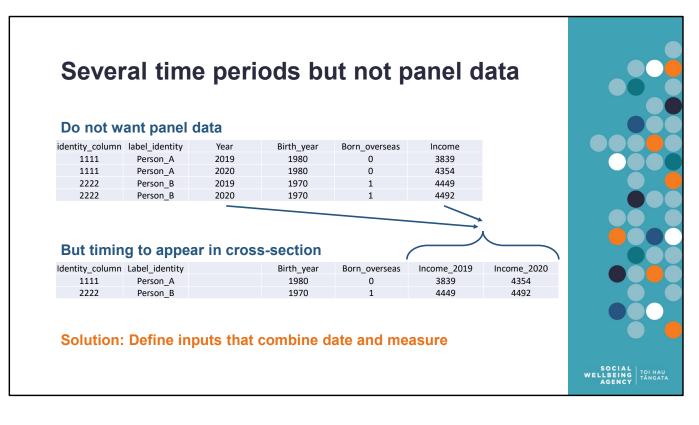
Producing panel date is best done by combining identity and summary period labels.

The Label\_identity and Label\_summary\_period inputs makes it easy to repeat the measures for different time-frames and different sub-populations.

For example, if you want to find total income for individuals in 2 set of populations (e.g. fathers and mothers) for 2019 and 2020, you can define your population with two identity labels – "mother" and "father" – and two summary labels – "2019" and "2020".

You can also use SQL columns for the label identity, start date, end date, and summary period label.

As an alternative to the example above: If you used a single table [defn\_res\_parents] that contained both mothers and fathers with two columns [snz\_uid] and [role], then the population file would only have four rows. Instead of "father" and "mother" in the Label\_identity column you would put [role] because the [role] column contains the labels father and mother.



Suppose you want a cross-section dataset but would like income for last year and this year. So rather than two rows per identity with columns [year] and [income] (as a panel dataset would have), you want one row per identity with columns [income\_this\_year] and [income\_last\_year].

Several approaches:

- 1. Assemble a panel and use lag/lead functions to fetch the value for the next year (but this duplicates all the other columns).
- 2. Assemble two datasets, a master dataset and a minimal dataset for the extra year then join them together (but can not do assembly in one step).
- 3. Define income for each year in the input measure and include both in the control file (recommended).

For our recommended approach, we can transform a measure table with columns [year] and [income] to a table with columns [year], [income], [income\_2019] and [income\_2020] by creating two new columns: [income\_2019] and [income\_2020]. In SQL this might look like:

```
SELECT *
    ,IIF([year] = 2019, [income], NULL) AS [income_2019]
    ,IIF([year] = 2020, [income], NULL) AS [income_2020]
FROM [db].[schema].[table]
```

# When all else fails, ask for help

#### **Gather error information**

Be well prepared

- What causes the error?
- What is the error message?
- What have you tried already?
- What is working?

#### **Request support**

• Poor preparation  $\rightarrow$  poor help

Read the speakers notes in this presentation - they give extra guidance



Some errors will be due to the assembly tool, and will not be fixable without a developer. The code is open source, so if you have a friendly R developer you could ask them. Otherwise you will need to ask for expert support.

Before you do, make sure you are well prepared.

- Work through this presentation, are there any details you have missed?
- Work through the guidance document
- Isolate the issue show how it occurs with only one measure and one population.
- Some staff in the Integrated Data team (access2microdata) are familiar with the tool

   check with them.

As a final resort, contact the tool developer / maintainer.

Remember, the developer is not a helpdesk, but a fellow researcher.

• How can you help me to help you?



Thank you for your time and attention.

I invite you to adopt the Dataset Assembly Tool for your IDI research. It has saved me a lot of time and headaches trying to build datasets manually. There is an initial library of definitions from my use of it. So I am confident it will add value to your work.